



# ANIMAL TRACKS

A newsletter for the Duke research community



January 2009

<http://vetmed.duhs.duke.edu>

## ACCLIMATIZATION/STABILIZATION OF ANIMALS

Acclimatizing and stabilizing newly received animals to their new environments is an important component of providing physiologically stable biologic models for research—especially during the stressful periods of animal transport such as winter. The Guide states that “newly received animals should be given a period for physiologic, psychologic and nutritional stabilization before their use.”

How can you maximize your research outcomes will minimizing effects of transport?

1. Order animals well in advance of when they will be required.
2. Assure all incoming animals have a minimum of 7 days acclimation to the facility prior to use.
3. If you have exemptions due to protocol requirements please advise DLAR husbandry staff.
4. Special husbandry and acclimation needs can be requested through DLAR husbandry staff to aide in acclimation of sensitive animals.
5. Animals under acclimatization or quarantine should not come in contact with animals already on study. Animals should be considered “infected” unless information confirms that the animals are disease free. In other words, all animals, even those you ‘think’ are clean, should NOT be considered clean until the surveillance tests confirm the animals ARE clean.

Acclimatization/stabilization is an important first step of great research and protects the integrity of the research outcomes!

## Upcoming Dates & Deadlines

January 5	New Protocol Deadline
January 9	Brown Bag Seminar: Performing the Required Protocol Lit Searches Protocol Requirements
January 12	Significant Change Deadline
February 2	New Protocol Deadline Significant Change Deadline
February 23	Significant Change Deadline Brown Bag Seminar: Preparing Your Lab for an IACUC Inspection
March 2	New Protocol deadline
March 2	Significant Change Deadline

## WHY WAIT? AVOID DELAYS!

Protocol Process Updates  
Obtaining OESO and EOHW Clearance  
for Animal Use Protocols & Amendments

There are two parallel processes at work with every protocol or amendment submitted to the Institutional Animal Care & Use Committee (IACUC). Both processes must be completed before the proposed work with animals can be accomplished. The processes are: 1) the IACUC must **APPROVE** the proposed activity; and 2) the Occupational Environmental Safety Office (OESO) and Employee Occupational Health and Wellness (EOHW) must **CLEAR** the proposed activity.

Since these processes are independent, it is possible the protocol will be approved by the IACUC many days prior to clearance being granted by OESO or EOHW. Many times protocol approval letters are held because one individual listed on the protocol who has not completed the web training, or one individual has not submitted a health questionnaire -- both very small but highly significant issues (and will delay your research)!

Did you know you can obtain OESO and EOHW clearance prior to submitting the protocol document for review? This will save time with the clearance, and assure that you can perform the requested as soon as the approval letter arrives!

What can you do?

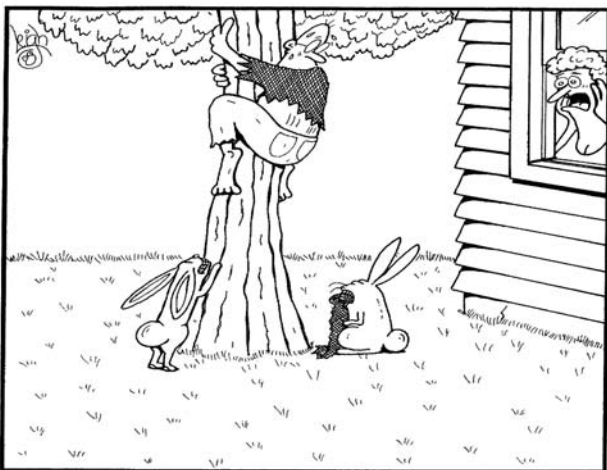
1. You can complete web training (or update annual requirements) via the on-line training (<http://www.safety.duke.edu>) and complete the Health Review for Animal Handlers (<https://www.hr.duke.edu/secure/eohw/animal.php>) at any time—there is no need to wait until the protocol is being processed.
2. Principle Investigators can check on the training status (OESO web training completed) of their laboratory staff. Contact Joan Catignani by email at ([catig001@mc.duke.edu](mailto:catig001@mc.duke.edu)) to establish the authority to review lab staff training records.

**OESO On-Line training modules:** All Duke Principle Investigators on animal protocols (whether handling animals or not) and all research associates handling animals must complete the on-line training modules Animal Handlers Part I and Animal Handler Part II located on the

## NOTIFICATION OF HAZARDOUS WORK IN ANIMAL CARE AREAS

When animal research involves the use of hazardous agents (i.e., infectious agents, hazardous chemicals, radiological agents, etc.) it is imperative (not to mention the right thing to do!) that workers be notified of potential risks and how to work safely when such risks are present. The role of the researcher is to ensure that hazardous agent information is provided to all research, animal care and occupational health staff. Here are some important highlights of working with hazardous agents at Duke:

- Assure all required signage is properly affixed and notification of pending (or ongoing) hazardous work has occurred prior to working with the agent in an animal use area.
- Prepare a Standard Operating Procedure (SOP) that outlines the safe work practices for the animal use area when hazardous agents are employed (must be OESO approved).
- Staff should be trained on proper use of PPE. Note: Personal Protective Equipment (PPE) is required but must be appropriate to the species and the hazardous agent being used.
- Notify DLAR via written communication, preferably e-mail, at least 2 business days prior to the use of hazardous agents in animals.
- Post hazard signs (available from OESO) in use areas. Information on the form must include; name of hazardous agent, building and room number, species in which agent will be used, emergency contact information for the PI and staff, copy of the SOP for safe handling of the agent being used.
- Notify a DLAR contact person to assure the above steps are completed:
  - Dr. Randall Reynolds ([randall.reynolds@duke.edu](mailto:randall.reynolds@duke.edu))
  - Peg Hogan ([hogan012@mc.duke.edu](mailto:hogan012@mc.duke.edu))
  - Dr. Francis Sun ([francis.sun@duke.edu](mailto:francis.sun@duke.edu))



"Good heavens, no wonder they're after you, Herb! They're herbivores!"

## ~ NEW PROGRAM POLICY ~

### PROTECTION OF PROTOCOL INFORMATION

The IACUC has approved a policy that establishes clear guidance on who may access your protocol information. Under this policy, animal program information is considered confidential, proprietary, or of a business nature and will be restricted to those with a bonafide 'need to know.' To review the full text policy, please visit the animal program website at <http://vetmed.duhs.duke.edu/> and select the link for 'POLICIES.'

The details of this new program policy are:

- All materials obtained from or sent to Duke researchers or their staff; from or to the IACUC; from or to the OAWA, and maintained in the program server as a master protocol or communication file of the Duke animal program are considered confidential and/or proprietary and/or protected business information.
- The researcher may share any or all of their protocol information with any individual they determine requires the document (e.g., laboratory staff, animal care team, granting agency, departmental staff).
- No member of the Duke animal care and use program may share animal protocol related information (e.g. files, reports, documents) with any individual, except under specific circumstances described in this policy.

#### Who has routine access to your protocol files?

- Certain program participants require access to your protocol information as a part of their job, these include: the IACUC leadership, OAWA staff, and DLAR veterinarians. IACUC members have access to your protocol information when they are performing their assigned duties as a member of the IACUC.
- Any individual listed as an 'approved protocol participant' in the ROLES section of your approved animal protocol, may receive a copy of the protocol file via email, upon request.
- Protocol information may be shared with any federal agency performing its obligations under statute or accreditation rules, such as the USDA, NIH, VAMC, or AAALAC).
- Specific requests for information will be approved for your Department Chairman; a Duke grants and contracts officer (e.g. ORA, ORS); or a Duke compliance official (e.g. SOM or DU Compliance Office).

In all cases, OAWA will alert you to the sharing of your protocol information except where there is a formal review of records concerning potential non-compliant activity. OAWA would be prohibited from communicating an external review of records..

This policy, while new in its existence, codifies the long standing animal program practices of protecting your research information.

Wishing you a productive research week!

## Ask Labby

~ Duke's 'Answer Critter' for Research Questions on Animal Use ~  
Send Questions to Labby at: IACUC@DUKE.EDU

**Dear Labby:** I am performing a surgical procedure in one of the DLAR surgical areas in the DLAR Vivarium. Can members of my surgical team take pictures of the surgical site during the procedure?

### Photofinish

*Dear Photofinish:* If you wish to take pictures within a Duke animal holding space, you must contact the Director of DLAR for approval.

←—————→  
**Dear Labby:** We are approved to perform toe clipping for identification as well as for genotyping. At what age can I perform this procedure?

### Shortcut

*Dear Shortcut:* Toe clipping in mice MUST occur prior to twelve (12) days of age, and only as approved by the IACUC. You state you are approved for toe clipping, but please remember approval is on a protocol-by-protocol basis and requires scientific justification. Call the OAWA for questions about obtaining approval.

←—————→  
**Dear Labby:** A DLAR veterinarian has recommended euthanasia for one of our mice. This is a valuable animal in our study and we don't want to lose it. Can we keep this animal in spite of the veterinarian's recommendations?

### Last One Standing

*Dear Ms. Standing:* As defined in the Veterinary Authority memo (located on the animal program web), DLAR veterinarians have the ultimate authority in determining the need for euthanasia and/or medical treatment in any research animal. If this animal is critically important to our research, then you should work with DLAR to develop a humane care plan or consider immediate embryo cryopreservation. If it becomes a humane issue, then the DLAR veterinary staff do have ultimate authority to determine if an animal must be euthanized.

←—————→  
**Dear Labby:** Can I use my personal vehicle to transport mice from the Vivarium to my laboratory across campus?

### Tommy Truckin'

*Dear Tommy:* Yes, you can use a person vehicle to transport animals *if* you have prior IACUC approval. This approval consists of an amendment to your protocol to include a vehicle for transportation. A personal animal transport vehicle must be IACUC inspected every 6 months. You can review the details of the transport of animals across campus see the policy located at: [http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy\\_on\\_animal\\_transport\\_around\\_campus.pdf](http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_animal_transport_around_campus.pdf)

## CONCORDANCE REVIEWS

A concordance review is a comparison between the grant submitted to the funding agency and the protocol reviewed by the IACCUC. Concordance reviews are required by many (but not all) granting agencies—especially federal ones. Generally, the granting agencies require an signed concordance letter PRIOR to release of the funds for the grant, so it is important to plan ahead!

Concordance reviews are performed by the OAWA/IACUC and require 5-7 business days to complete. Why so long? Because a concordance review compares each and every procedure stated in the grant with the procedures approved in the protocol, and they must match! To be concordant, everything you told your granting agency you would do with animals, even if it will not be done until the 4th or 5th year, must be IACUC approved before the concordance letter can be generated.

The OAWA/IACUC requires the whole grant EXCEPT the biosketches and the budget pages. It is preferred that an electronic copy be provided when the protocol is submitted (if you have the grant at that time). If not, a concordance review can be done at any time, but it will take the 5-7 business days to complete.

The secret? PLAN AHEAD! If you think you MAY need a concordance review, request it up front, because these reviews cannot be rushed, and do require time to complete.



Why Wait? .... From Page 1

OESO web site. Log in with your Duke Net Id and password. The On-Line training tab is located on the left hand side of the main page. To access the Animal Handlers courses click on Courses Available on Line. The two components are Regulations Impacting Animal Care and Use and Veterinary Care. There is a quiz for each module. Once completed the data is maintained by OESO and can be accessed by animal program staff during the protocol review process. These modules are required only once (no annual update on these modules).

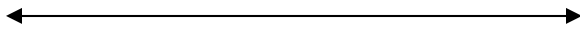
EOHW clearance: All research staff who will work with animal models must complete the "Health Review for Animal Handlers" on the Duke EOHW web site [www.hr.duke.edu/eohw](http://www.hr.duke.edu/eohw). The link is located on the main page. Click on Health Review for Animal Handlers. You will need a Duke unique ID in order to complete the form. Fill out all required information and click on submit when finished. You do not need to submit a hard copy. Questionnaires will be reviewed by EOHW medical staff. If there are any concerns with respect to occupational risks you will be contacted by one of the staff nurses. If you have additional questions after submission you may contact the EOHW office at 6864-3136 and select option #2.

If you need additional assistance with this process please call Bill Wade at 668.6722 in the OAWA office.

**Dear Labby:** We are approved to perform tail clipping for genotyping. Do I have to use anesthesia in order to perform this technique?

**Dr. Gene O'Type**

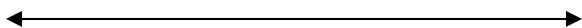
**Dear Dr. Type:** According to the policy on tail clipping, anesthesia is not required if tail clipping is performed *prior* to twenty-one (21) days of age. After 21 days of age (generally this is the date of weaning), anesthesia and analgesia are required. If anesthesia cannot be provided to a post-21 day animal, scientific justification **MUST** be pre-approved by the IACUC.



**Dear Labby:** Recently I had an amendment reviewed and approved by the IACUC and it took 12 days. They said it was a 'significant' amendment. If I knew what makes a proposed protocol amendment 'significant', I might be able to plan my experiments better. What makes amendments 'significant'?

**Siggy**

**Dear Siggy:** A 'significant' amendment usually involves things such as changes in species, an increase greater than 20% of approved animal numbers, housing for greater than twelve (12) hours in a non-DLAR managed housing facility, non survival to survival surgery, increased invasiveness of a procedure, increased proportion of animal deaths, increase in duration of pain, discomfort, or distress to an animal, administration of a hazardous agent, change in principal investigator, addition of neuromuscular blocking agents, or a change in euthanasia procedure for AVMA conditional or not acceptable procedures). It is not possible to list all potential significant amendments, so please review the amendment policy on the animal program web or call the OAWA to ask if what you are proposing will be considered significant.



**Dear Labby:** What are the minimal requirements for aseptic surgery in rodents?

**Dr. Stiches**

**Dear Doc:** The minimum aseptic technique for survival surgery in rodents includes: sterile instruments and supplies (initial sterilization with autoclave or gas and use of a glass bead sterilizer or other IACUC approved method between animals), face mask and sterile gloves. A surgical gown and/or surgical scrub are suggested. An area for rodent surgery should be a dedicated space in the laboratory. Surgical site preparation must include clipping of fur, surgical scrub and disinfection of the surgical site (a '3 layered' process). You can review the details in the IACUC policy on the animal program web site.

## Animal rights extremists guilty of campaign to blackmail Huntingdon Life Sciences

From: the London Times

Seven animal rights extremists who subjected scores of victims to a "climate of fear" have been convicted of coordinating a six-year campaign of blackmail and menaces to shut down Huntingdon Life Sciences. The three women and four men were found guilty after a two-year, £3.5 million police investigation into Stop Huntingdon Animal Cruelty, an international campaign to close down the Cambridgeshire-based animal research laboratory.

Prosecutors believe that among these members SHAC's hierarchy were some of the key figures in the Animal Liberation Front, the often violent movement that acts as an umbrella for much animal rights extremism worldwide.

Two of SHAC's founding members, Gregg Avery, 41, and his wife Natasha, 39, along with fellow activist Daniel Amos, 22, pleaded guilty earlier this year to conspiracy to blackmail. Following a three-month trial at Winchester Crown Court, Avery's former wife and fellow SHAC founder Heather Nicholson, 41, was found guilty of the same offence. Three further conspirators, Daniel Wadham, 21, Gerrah Selby, 20 and Gavin Medd-Hall, 45 were also convicted. All are expected to be sentenced next month. The maximum sentence for conspiracy to blackmail is 14 years imprisonment. An eighth defendant, 51-year-old Trevor Holmes from Newcastle, was acquitted.

One of the jurors refused to be seen in court while the verdict was announced today after 33 hours and 48 minutes of deliberation. Sentencing will take place on January 19 at Winchester Crown Court. Selby, Wadham and Medd-Hall were released on conditional bail, while Nicholson was remanded in custody.

SHAC's victims received threatening letters, hoax bombs and sanitary towels allegedly contaminated with the HIV virus, while their neighbours were sent anonymous letters warning them that they lived close to a paedophile. The managing director of one targeted company received a letter in December 2006 that threatened: "We will attack your property, your family or you, whichever we see fit . . . The screams of the animals are in our heads. We will not fail them. You will pay for their agony."

Nocturnal "home visits" from extremists left cars covered in acid, menacing messages painted on houses and ALF slogans daubed on nearby roads. The targets, who worked for companies that did business directly or indirectly with HLS, were usually targeted after they were listed as 'collaborators' on SHAC's website. This persecution, often of people who were only loosely connected with animal research, normally stopped only when a victim's company agreed to sever its links with the laboratory. Avery and the rest of his core group denied any links to the ALF or the blackmailing of HLS "collaborators" whose details appeared on SHAC's website.

**If you have a research question for Labby, just send it to Dear Labby at the email address [IACUC@Duke.edu](mailto:IACUC@Duke.edu). Then watch for the next issue of Animal Tracks for the response!**



## OAWA's Brown Bag Seminar

Friday, January 9<sup>th</sup>, 2009

Noon – 1 p.m.

Bryan Research Building: Room 103

Mr. Teddy Gray  
Head, Biological and  
Environmental Sciences Library

Will be presenting:

# Alternatives Searching for the Researcher

**THIS SEMINAR WILL COVER HOW TO EFFECTIVELY SEARCH FOR THE THREE R'S (REFINEMENT, REDUCTION, REPLACEMENT) OF ANIMAL RESEARCH. WE WILL HIGHLIGHT THE BEST LITERATURE DATABASES TO SEARCH AND HOW TO CHOOSE AMONG THEM. WE WILL THEN COVER CREATING EFFECTIVE SEARCH STRATEGIES INCLUDING THE USE OF SUBJECT HEADINGS AND DEMONSTRATE SAMPLE SEARCHES IN PUBMED. FINALLY, WE WILL POINT OUT ADDITIONAL RESOURCES FOR RESEARCHERS USING ANIMAL MODELS.**

The presentation will be on **Friday, January 9<sup>th</sup>, 2009 from Noon to 1 p.m.**

The session will be held in room **103 of the Bryan Research Building**, located at 421 Research Drive, on Duke University's West Campus.

Attendees are encouraged to bring a lunch. OAWA will provide drinks and desserts.

Please plan on arriving prior to noon in order to get refreshments, sign in, and be seated.

For those who will be coming from off campus, driving directions and parking information can be found at the following link: <http://neuro.duke.edu/Links/map.htm>

**This session will count for 1 CEU of AALAS In-house Training Credit**