



ANIMAL TRACKS



A newsletter for the Duke research community

April 2008

<http://vetmed.duhs.duke.edu>

Animal Program Policies

Did you know the Duke animal program has various policies in place that affect use of animals at Duke? All policies are posted at the animal program web site. There are three types of program policies: **Animal Use, Protocol Management** (IACUC Processes), and **Program Management and Monitoring**. There is also a section under the index of program policies located on Duke Animal Care and Use Program website which lists aspects of controlled substance use in the animal care program. This issue of *Animal Tracks* is devoted to Duke animal program policies. We will showcase some of the more important and more commonly used policies, but invite your attention to the web site for a full discussion.

Protocol Management (IACUC Process) Policies:

These IACUC policies deal with issues related to animal use protocols.

- Amendments (Minor or Significant) to Approved Protocols
- Administrative Termination of Protocols
- Administrative Extension of Protocols
- Deferred Annual Review and Animal Use
- Faculty Protocol Requirement for Work Performed Off Site (collaborative)
- Protocol Distribution
- Required Signatures (OESO, EOHW & Others)
- Significant Amendment to Approved Protocols
- Annual Protocol Approval and CO2 Training Requirements
- Use of the DLAR Holding Protocol
- Use of Telecommunications for Conducting IACUC Business
- Working with Animals at Other Institutions (No Protocol Required)

Animal Use Policies: These IACUC policies deal with issues, procedures, and techniques concerning the use of animals at Duke University.

- Adoption of Animals Post-Research
- Acclimatization of Research Animals Prior to Use
- Aseptic Technique
- Alcohol as a Disinfectant (Surgery)
- Cage Density Requirements (Mice)
- Clinical Conditions Warranting Intensive Care
- Euthanasia (Approved Methods for All Species)
- Euthanasia (Mice Fetuses & Neonates)
- Frog Oocyte Collection
- Importing Animals or Tissues into Duke
- Medical Record Keeping
- Notification of Hazardous Work Using Animals
- OESO and Employee Health Requirements for Animal Handlers
- OESO Approval of Off Campus Animal Activities
- Principal Investigator Definition
- Protective Clothing Requirements
- Q Fever (Managing the Risks of Working With Ungulates)
- Source of Canines for Duke Related Research
- Surgical Disinfectants
- Transfer of Post Surgical Animals (Investigator to DLAR)
- Transport of Animals Around Campus
- Transport of Animals Through Patient Care Areas (Duke North / Duke South / Clinics)
- Transport of Animals by DLAR
- Tail Clipping of Mice for Tissue Collection or Identification
- Top Shelf Housing of Rodents
- Training Requirements for Animal Care Staff
- Tumor Burden in Rodents (Maximum)
- Vaporizer Maintenance and Calibration

Program (Management and Monitoring) Policies:

These policies are concerned with program wide policies of management and monitoring

- Compliance Liaison Program
- Controlled Substance Management
- Animal Program's Code of Ethics
- Emergency Responsibilities and Action
- Photography Within Duke Vivaria (Animal Holding/Use Areas)
- PHS Policy on CO2 and Secondary Euthanasia Techniques
- PHS Published Guidance and Policies
- US Government Principles
- Veterinary Authority
- Vivarium Work House
- Vivarium Access

Upcoming Dates & Deadlines

April 1	New protocol deadline
April 3	SC meeting
April 7	SC amendment deadline
April 17	SC meeting
April 21	SC amendment deadline
April 21	Brown Bag Seminar: Aseptic Rodent Surgery at Duke
April 24	IACUC meeting
May 1	SC meeting
May 5	New protocol deadline
May 5	SC amendment deadline



POLICY Acclimatization / Stabilization of Animals

This policy is in place to provide physiologically stable biologic models for research. The *Guide for the Care and Use of Laboratory Animals* states "newly received animals should be given a period for physiologic, psychologic, and nutritional stabilization before their use." For all newly received animals the IACUC generally requires a minimum of a forty-eight (48) hour acclimation period before any experimental use can occur. Exceptions to the 48-hour acclimation period requires a written justification included in the protocol. To review a complete copy of this policy, visit this link:
http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_acclimatization.pdf

POLICY:

Cage Space Requirements for Mice

Mice are maintained at a density that is consistent with the *Guide for the Care and Use of Laboratory Animals*. It is the Principal Investigator's responsibility to monitor animal housing density at a frequency sufficient to allow proper weaning (21 days of age unless an exemption is granted by the IACUC for delayed weaning) and adherence to the standards of this IACUC policy. When deviations from the standards outlined in this policy are identified, the research staff should correct the deficiencies immediately. Cages in which DLAR has identified an overcrowded housing situation must be separated by laboratory personnel within twenty-four (24) hours of DLAR notification.

Overcrowded housing conditions can be prevented by implementing procedures such as removing the male from a harem breeding scheme once breeding has occurred and by housing females singly up to one week prior to gestation. A recent change to this policy has determined single housing of pregnant females does not constitute an exemption to the *Guide*, thus separate IACUC approval is not necessary. To review this IACUC policy, go to:
<http://vetmed.duhs.duke.edu/documents/iacuc/pdf/Policy%20on%20Cage%20Density%20Requirements.pdf>

POLICY: Principal Investigators

The PI is responsible for all animal work performed under his or her approved animal use protocol or SOP. The PI must complete all required animal handling training, assure sufficient training for all individuals working under his or her protocols, and report any unexpected or adverse events that affect animals under his or her charge. The PI must also provide requires signatures and assurances for all IACUC documents prior to the approval of those documents.

The definition of a PI at Duke is one of the following: faculty member of Duke University or Duke University Medical Center (DUMC); veterinarian on staff at DLAR, DLC, or VAMC; or any Duke or DUMC staff member, student, or third party individual with a faculty member or a veterinarian serving as a sponsor for the proposed activity.

To review this policy, go to:
http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_definition_of_a_principal_investigator.pdf

Policy Q&A

Question: I am performing a surgical procedure in one of the DLAR surgical areas in the DLAR Vivarium. Can members of my surgical team take pictures of the surgical site during the procedure?

Answer: If you wish to take pictures within a DLAR managed housing space, you must contact the Director of DLAR. See the policy on photography located at:
http://vetmed.duhs.duke.edu/policy_on_photography.htm

Question: We are approved to perform toe clipping for identification as well as for genotyping. At what age can I perform this procedure?

Answer: Toe clipping in mice MUST occur prior to twelve (12) days of age. See the related policy at:
http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_tail_and_toe_clipping_of_mice.pdf

Question: A DLAR veterinarian has recommended euthanasia for one of our mice. This is a valuable animal in our study. Can we keep this animal in spite of the veterinarian's recommendations?

Answer: No. DLAR veterinarians have the ultimate authority in determining the need for euthanasia and/or medical treatment in any research animal. See the policy on veterinary authority located at:
http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_veterinary_authority.pdf

Question: Can I use my personal vehicle to transport mice from the Vivarium to my laboratory across campus?

Answer: Yes, you can use a person vehicle to transport animals *if* you have prior IACUC approval. This approval consists of an amendment to your protocol to include a vehicle for transportation and the successful completion of an IACUC inspection of your vehicle every six months. For more information about transport of animals across campus see the policy located at:
http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_animal_transport_around_campus.pdf

Question: We are approved to perform tail clipping for genotyping. Do I have to use anesthesia in order to perform this technique?

Answer: According to the policy on tail clipping, anesthesia is not required if tail clipping is performed *prior* to twenty-one (21) days of age. See the policy on tail and toe clipping located at:
http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_tail_and_toe_clipping_of_mice.pdf

Question: What makes a proposed protocol amendment 'significant'?

Answer: A significant, or major, amendment is required when there are changes in species, an increase greater than 20% of approved animal numbers, housing for greater than twelve (12) hours in a non-DLAR managed housing facility, non survival to survival surgery, increased invasiveness of a procedure, increased proportion of animal deaths, increase in duration of pain, discomfort, or distress to an animal, administration of a hazardous agent, change in principal investigator, addition of neuromuscular blocking agents, or a change in euthanasia procedure for AVMA conditional or not acceptable procedures). For more information about protocol amendments, please review the policy located at:
http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_IACUC_review_and_approval_protocol_amendments.pdf

Question: What are the minimal requirements for aseptic surgery in rodents?

Answer: The minimum aseptic technique for survival surgery in rodents includes: sterile instruments and supplies (initial sterilization with autoclave or gas and use of a glass bead sterilizer or other IACUC approved method between animals), face mask and sterile gloves, gown and/or surgical scrub is suggested, area for rodent surgery should be a dedicated space in the laboratory, and surgical site preparation including clipping of fur, surgical scrub and disinfection of the surgical site (recommend betadine alternated with alcohol three (3) times). For more information view the policy at:
http://vetmed.duhs.duke.edu/documents/iacuc/pdf/policy_on_aseptic_technique.pdf

OAWA's Brown Bag Seminar

Monday, April 21st, 2008

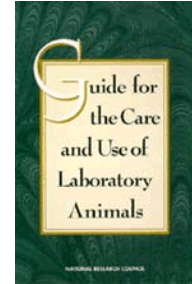
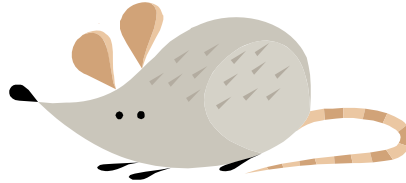
Noon – 1 p.m.

Bryan Research Building: Room 103

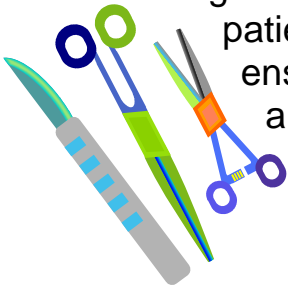
**Dr. Ron Banks, OAWA Director
Will be presenting:**



Rodent Survival Surgery: How to Maintain Proper Aseptic Technique



According to *The Guide for the Care and Use of Laboratory Animals* and Duke's Policy on Aseptic Technique, all survival surgeries should follow the same principles of aseptic technique. There are many aspects to aseptic technique that need to be addressed in order to maintain sterility during procedures. You must consider the site of the procedure, the instruments to be used, the patient, and the surgeon when preparing for a procedure. The specific areas of proper preparation of the surgical location, acceptable instrument sterilization methods, proper patient skin preparation, and proper steps the surgeon should take to ensure and maintain sterility throughout the procedure and between animals will be covered.



The presentation will be on **April 21st, 2008** in room **103 of the Bryan Research Building**, located at 421 Research Drive, on Duke University's West Campus.

OAWA will provide drinks and desserts. The session will begin promptly at Noon. Please arrive early to sign-in and find a seat.

For those who will be coming from off campus, driving directions and parking information can be found at the following link: <http://neuro.duke.edu/Links/map.htm>

This session will count for 1 CEU of AALAS In-house Training Credit